

Coronavirus: A Possible Cause of Reduced Male Fertility

Running title: Coronavirus and male fertility

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Abstract

In lately December 2019, a novel coronavirus (SARS-CoV-2) outbreak occurred in Wuhan, PR China. It is a high contagious virus that has threatened human health worldwide.

SARS-CoV-2 infection, termed COVID-19, causes rapidly developing lung lesions that can lead to multiple organ failure in a short period. Whenever a novel virus emerges, reproductive risk assessments should be performed after infection. In this review, we show that male

fertility might be damaged by coronavirus associated with (i) direct cytopathic effects derived

from viral replication and viral dissemination in the testis; and (ii) indirect damage to male

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fertility derived from immunopathology. In this review, we briefly describe the impaired fertility of humans and animals infected with coronaviruses to deduce the impact of the new coronavirus on male fertility. Together with information related to other coronaviruses, we extrapolate this knowledge to the new coronavirus SARS-CoV-2, which may have a significant impact on our understanding of the pathophysiology of this new virus.

Keywords: Coronavirus, Fertility, Male, SARS-CoV-2, Testis

1. Introduction

Coronaviruses are the largest family of positive-stranded RNA viruses, which includes 30 members at present. They are widely distributed in nature, including infections of humans and other mammals. In recent years, new coronaviruses have caused problems worldwide in cycles, such as severe acute respiratory syndrome coronavirus (SARS-CoV) occurring in 2002, and Middle East respiratory syndrome coronavirus (MERS-CoV) being first identified in 2012. In 2019, a new highly contagious virus broke out in Wuhan, Hubei province, China, termed SARS-CoV-2, representing the seventh member of enveloped RNA coronaviruses¹. The 2019 novel coronavirus disease (COVID-19) caused by SARS-CoV-2 has common clinical manifestations such as fever, dry cough, and in severe cases, multiple organ damage²⁻⁴.

Regarding the critical molecule for SARS-CoV-2 transmission, the receptor angiotensin I converting Enzyme 2 (ACE2) for virus cell entry and transmembrane serine protease 2 TMPRSS2 for priming the S protein⁵, are co-expressed in the testis and male genital tract⁶, which suggests a high possibility that the virus targets the testis and male genital tract during

infection. It was reported that over 25 viruses could enter human semen and negatively affect sperm or male fertility⁷, such as HSV⁸ and HIV⁹. Whether SARS-CoV-2 may have the same the effect on males is an important question that was not answered unambiguously in a preliminary investigation¹⁰.

To date, studies¹¹⁻¹³ have confirmed the absence of SARS-CoV-2 RNA in the semen of patients with COVID-19. Conversely, The results of Li et al.'s study were inconsistent with those of previous studies and detected of SARS-CoV-2 in 6 of 38 semen samples¹⁴. Similarly, Yang et al. reported that one case (1/12) with a high viral load was positive for viral RNA after post-mortem examinations of testicular tissue¹⁵, which supported the idea that high viral loads in patients with severe disease symptoms might reach the threshold to cross the blood-testis barrier¹⁶. On the other hand, a study showed that compared with patients with mild disease, patients with severe COVID-19 have significantly lower testosterone levels¹⁷, suggesting that the co-expression of ACE2 and TMPRSS2 on Leydig cells might make them susceptible to SARS-CoV-2 infection and thus compromise testosterone secretion¹⁸. However, considering the high false-negative results for SARS-CoV-2 using RT-PCR¹⁹, as well as the limitation of the small sample size and selection bias mostly obtained from recovering mild cases¹⁰, we still need to be cautious when evaluating this data. Nevertheless, it is well known that coronaviruses can contribute to high morbidity and mortality in both humans and animals^{20,21}. A study has demonstrated orchitis in patients with SARS, with detrimental effects in the testis, suggesting that coronavirus can infect the male reproductive tract and impair male reproduction²². SARS-CoV-2 and SARS-CoV share some common clinical manifestations, which supports the hypothesis that the new coronavirus might directly infect the testes and

male reproductive system. Therefore, we should be vigilant about the impact on male reproduction in patients with COVID-19. In addition, the blood-testis barrier might allow the testes to act as a special reservoir to protect viruses against antiviral agents²³, which is a key reason for considering the testes as a particularly important organ for study in the context of the SARS-CoV-2 pandemic, and it is especially important because the coronavirus family has been identified the culprit causing orchitis in both humans (SARS-CoV)²² and animals (feline coronavirus and avian coronavirus)^{24,25}. Using evidence from previous studies of coronavirus-infected animals and humans, the implications of this review may help us to understand the impact of SARS-CoV-2 on male reproductive capacity.

2. Direct virus-induced cytopathic effects

Various viruses can replicate in the male reproductive tract, such as HEV²⁶ and ZIKV^{27,28}, which eventually lead to testicular atrophy and male infertility. Viral infection of the male genital tract can provide insights into possible male fertility impairment after SARS-CoV-2 infection. SARS-CoV-2 enters cells by binding ACE2 and via priming by TMPRSS2. ACE2 is a membrane-associated secretase that is expressed primarily on endothelial cells and is the host cell receptor for SARS and SARS-CoV-2²⁹⁻³¹. Notably, ACE2 is highly tissue-specific, with significant levels being detected only in the heart, kidneys, testes, and gastrointestinal tract³²⁻³⁴. In the testes, ACE2 is expressed only in spermatogenic cells and testis somatic cells, suggesting a high potential for testicular damage and spermatogenesis disruption when the virus combines with this metalloprotease³⁵. TMPRSS2, as an essential protease for viral infection, is highly expressed in spermatogonia and spermatids¹⁸. The co-expression of ACE2 and TMPRSS2 in spermatogonia and Leydig cells

implied that the testis might be a high-risk organ that is vulnerable to SARS-CoV-2 infection, which might result in testicular degeneration and male infertility. SARS coronaviruses, whose expressed proteins share 76% amino acid sequence identity with those of SARS-CoV-2, were detected in testis somatic cells³⁶. This observation supports the hypothesis that the SARS-CoV-2 might concentrate on testis cells to dysregulate their function.

2.1 Direct virus-induced damage of the testis

Viral replication in cells contributes directly to microscopy-detected lesions, which eventually result in spermatogonia necrosis²⁶, such as in a ram model of infection by Bluetongue virus (BTV) (an arbovirus of ruminants), which showed testicular parenchyma damage and the destruction of the Sertoli cells caused by viral replication-induced cytopathic effects³⁷. Coronaviruses might use a similar mechanism in humans to impair male fertility. ACE2 is the crucial determinant of coronavirus infection, tissue tropism, and subsequent viral replication^{38,39}. The expression pattern of ACE2 in adult human testis at the level of single-cell transcriptomes was shown to be predominantly enriched in Leydig and Sertoli cells⁶. Besides, alternative receptor Basigin (BSG) and protease Cathepsin L (CTSL) were also detected in Leydig cells⁴⁰, which can mediate SARS-CoV-2 into cells. Data from autopsies of 12 patients with COVID-19 showed a dramatic reduction in Leydig cells in the interstitium¹⁵, supporting the speculation that SARS-CoV-2 could display tropism for Leydig cells, ultimately leading to ultrastructural lesions and decreased numbers of Leydig cells. Leydig cells occur in clusters between blood vessels and seminiferous tubules, producing the vast majority of androgens in men⁴¹. The replication of SARS-CoV-2 in testosterone-producing Leydig cells might disrupt testosterone production. Indeed, a recent

study confirmed that patients with COVID-19 suffered hypogonadotropic hypogonadism as
the disease the progressed, implying that the secretory function of Leydig cells might be
impaired by the novel coronavirus¹⁷. Testosterone is essential to preserve male fertility and to
support Sertoli cell maturation and the development of Leydig cells⁴². Extensive evidence
from clinical and laboratory studies implied that testosterone deficiency is accompanied by
atrophy of the testicular parenchyma and degradation in the seminiferous tubules^{43,44}, in
summary, testosterone is necessary for men to maintain the blood-testis barrier,
spermatogenesis, and fertility. Alterations in male sex hormone levels induced by
SARS-CoV-2 might negatively affect male reproduction. Therefore, special attention should
be paid to andrology examinations and hormone assessments on men recovering from
COVID-19, as well as exploring the possible long-term outcomes of SARS-CoV-2 infection.

Sertoli cells are the only somatic cells in the tubules that directly contact with
spermatogenic cells, and control the differentiation of spermatogenic cells via paracrine
signals⁴⁵. Inhibin B is secreted by Sertoli cells, and compared with follicle-stimulating
hormone (FSH) or luteinizing hormone (LH), it is an ideal marker for spermatogenesis and a
better indicator of sterility^{46,47}. SARS-CoV-2 has a high affinity for human ACE2, which
suggests that the virus might concentrate on Sertoli cells. Indeed, inhibin B levels decreased
after hepatitis E virus infection in mice, which was attributed to damage of the Sertoli cells in
the testes²⁶. Accumulating evidence suggests that the coronavirus family has an affinity for
these testes cells, for example, Avian infectious bronchitis virus (IBV), a subtype of
coronavirus, can cause acute respiratory infections in birds⁴⁸, and was detected in Sertoli cells
of the testes of infected roosters using immunofluorescence⁴⁹. Roosters vaccinated with live

attenuated IBV showed significantly reduced serum androgen concentrations compared with non-vaccinated roosters and could cause infertility in roosters⁵⁰. Considering that IBV causes a similar severe acute respiratory syndrome to SARS-CoV-2, we hypothesized that the same mechanism might be used by SARS-CoV-2 to spread in Sertoli cells. In addition, roosters vaccinated with live attenuated IBV might provide an animal model of how SARS-CoV-2 replicates in cells and causes pathogenic effects in the testis.

Additionally, coronavirus might directly disrupt the microenvironment of the testis that supports spermatogenesis. In CoV-infected roosters, histological analysis revealed the disruption of seminiferous tubules and loss of the basement membrane, leading to the destruction of the spermatogenesis microenvironment, which contributed to the reduction of the live sperm concentration⁵¹. Thus, testicular degeneration is possibly the result of several overlapping factors once a coronavirus infects the male genital tract, and this might also be the case for SARS-CoV-2.

2.2 Virus-induced damage of spermatogenesis directly

Viruses can be detected in semen directly. SARS-CoV-2 RNA has been isolated from rectal swabs and respiratory tract swabs⁵². Currently, the question of whether the virus can infect semen needs an answer. According to a recent study of scRNA-seq data in adult human testes, ACE2 and TMPRSS2 are highly co-expressed in spermatogonia, which are enriched in the gene ontology (GO) categories relating to viral reproduction and transmission⁶. Therefore, it is reasonable to hypothesize that there is a high risk of SARS-CoV-2 presence in seminal fluid⁵³. However, a few case reports have investigated this issue, and the presence of SARS-COV-2 in semen remains ambiguous¹¹⁻¹⁴. Notably, gene ontology (GO) enrichment

analysis illustrated that cell junction and immunity-related GO terms were enriched in ACE2-positive Leydig/Sertoli cells; therefore, cell-cell junctions might allow the transfer of SARS-CoV-2⁶, which might represent one explanation of the highly contagious nature of this novel coronavirus and could have implications for sexual and reproductive behavior⁵⁴. Taken together, there is a critical need to verify virus infection semen and whether sexual transmission of SARS-CoV-2 can indeed occur.

With regard to research on coronavirus-infection animals, IBV has been isolated from testicles and semen in roosters ^{51,55}, and when insemination using IBV-spiked semen was performed, IBV RNA could be detected in all the hens, and the weight of eggs laid by the hens inseminated with IBV-spiked semen was significantly reduced ⁵⁵. Knowledge of other coronaviruses present in semen might encourages researchers to look at semen and sexual transmission, to determine whether SARS-CoV-2 can be sexually transmitted like IBV; however, the results will need to be cautiously interpreted. Nevertheless, we should remain vigilant to this possibility, which has important implications in reproductive medicine, especially viral transmission facilitated by ART, such as intracytoplasmic sperm injection (ICSI)⁵⁶, sperm cryopreservation, and the prevention of transmission. During this epidemic, sperm cryobank must introduce precautionary measures: first, we recommend that semen from SARS-CoV-2-positive men is cryopreserved in a highly secure, separate container, such as a vapor cryostorage tank. Secondly, all donors must undergo mandatory SARS-CoV-2 testing. Thirdly, abstinence or condom use might be considered as preventive measure for patients with COVID-19.

Virus binding to ACE2-expressing spermatogonia would disrupt spermatogenesis⁶.

Rooster vaccination with coronavirus caused a significant reduction in daily sperm production^{25,57}. A recent report also confirmed that semen quality parameters were impaired in patients with moderate infection of COVID-19⁵⁸. Hence, the risk of SARS-CoV-2 virus infection to semen parameters may not be negligible. Notably, the long-term impact on semen parameters of SARS-CoV-2 infection, as well as semen examination, is required during follow-up patients recovering from COVID-19, especially men who plan to have children.

2.3 Direct virus-induced damage of the epididymis

In animal models of coronavirus infection, one of the major clinical symptoms is epididymal stone formation^{51,59,60}. IBV replication in roosters' testes might result in severe cellular micropathological damage, which in the long-term can lead to the presence of epididymal stones. The presence of stones is associated with reduced fertility and adverse effects on sperm function⁶¹, eventually resulting in the collapse of the seminiferous tubules and cessation of spermatogenesis. The epididymis is a crucial region for sperm maturation, which is pivotal for sperm to obtain the motile ability and fertile capacity. Dysfunction in this area can compromise sperm maturation and further impair sperm quality, such as decreased sperm motility, increased DNA damage, changed membrane lipids, and the acrosome reaction⁶². If the behavior of coronavirus infection in humans is similar to that in animal models, we should pay attention to the epididymis to protect it from SARS-CoV-2-induced destruction.

In view of these result, we suggest prompting a comprehensive genitourinary examination for patients with COVID-19, including alterations in semen parameters, such as the acrosome reaction, DNA damage, and sperm motility.

3. Indirect immune-mediated damage to male fertility

The testicle is an immunologically privileged organ, the blood-testis barrier (BTB) protects the testes against pathogen invasion⁶³. In healthy fertile men, various immune cells and cytokines produced by non-immune cells are indispensable to ensure male fertility⁶⁴, in which they maintain the testicular microenvironment balance and male reproductive health within the intricate and active environment of the seminiferous epithelium. Testicular tissue development benefits from immune cells and their cytokines, and the immune response is critical to control and eliminate viral infection⁶⁵. Cytokines are important for the immune response to viral infections by regulating the expansion and location of leukocytes. However, infection and inflammation might disrupt the immune balance in the body, either through immune insufficiency or overactivation, possibly leading to devastating effects in humans⁶⁶. Immune pathology associated with an uncontrolled immune response might give rise to testicular parenchyma destruction when the BTB is damaged by virus infection⁶⁷, and any associated functional impairment could lead to male infertility.

3.1 Cytokine-mediated infertility

SARS-CoV-2 has proven effects on multiple organs throughout the body⁶⁸, accompanied by immunopathological reactions and high cytokine storms. In the plasma of patients with COVID-19 in intensive care units, higher plasma levels of cytokines were detected, implying that a cytokine storm might aggravate the infection in patients with COVID-19^{2,3}. Actually, this coincided with the research that patients with COVID-19 presented a typical profile of hyper inflammation, such as TNF- α , IL-6, and IL-1 β ⁶⁹. Cytokines are beneficial to testicular function and sperm production, as well as testicular

immunity privilege⁷⁰⁻⁷². However, a high concentration of inflammatory cytokines could contribute to the progression of sexual dysfunction⁷³. Thus, a change in cytokine production can lead to fertility problems^{66,74}. Cytokine-mediated suppression of the hypothalamic-pituitary-testicular axis could lead to a decrease in serum testosterone, such as IL1 leading to inactivation of the P450/c17 lyase that converts progestins into androgens in immunopathogenesis, which will result in decreased testosterone and sperm production^{60,75-77}. This corroborated the results showing a dramatic decline serum testosterone in 17 patients with severe COVID-19, which might even predict poor progression of COVID-19 infection⁷⁸. With a history of COVID-19 disease, SARS-CoV-2 infection can attribute to male hypogonadism⁷⁹, thus it is recommended to measure testosterone levels when a patient is detected as positive for SARS-CoV-2 RNA and conduct appropriate testosterone treatment if necessary. Studies detected dramatic increases in IL6 levels in patients with COVID-19^{80,81}. Immunopathologically, high IL6 expression correlates with a systemic inflammatory milieu that disrupts the integrity of the blood-testis barrier⁸². As a result of blood-testis dissemination, the virus might damage testicular tissue directly. Furthermore, COVID-19-induced changes to the cytokine microenvironment might even lead to testicular cancer⁶⁴, which could have long-term adverse effects on the recovery of patients, and represents a second long-term matter of concern. Hence, it should be noted that the cytokine storm introduced by SARS-CoV-2 could be associated with immunopathogenesis, which might contribute to testicular dysfunction and reduced male fertility. Nevertheless, this hypothesis requires follow-up confirmation, and the exploration of possible short- and long-term consequences on their andrological health.

3.2 Inflammation-mediated infertility

The blood-testicular barrier might not be a perfect barrier to viruses under systemic or local inflammation⁷. To eliminate the virus infection, an inflammatory cytokines storm can recruit leukocytes, resulting in inflammation characterized by leukocyte infiltration in the interstitial tissue of the testes, which, as a feature of human testicular orchitis, might lead to male infertility. Actually, there is a high risk of that men with SARS-CoV-2 might suffer from an orchitis-like syndrome³⁵. Pan et al. confirmed that six patients (19%) with COVID-19 suffered from orchitis ¹¹. Recently, a study of 12 deceased patients with COVID-19 also revealed viral orchitis characteristics, with T lymphocyte intrusion into the testicular parenchyma, accompanied by significant seminiferous tubular injury¹⁵. Interestingly, the histopathological features of the testes in patients with SARS also overlap with those in patients with COVID-19: All testes being full of leukocyte infiltration and wide-ranging germ cell deterioration, with thickened basement membranes ⁸³, which supports the hypothesis that the coronavirus-induced adaptive immune response might play a vital role in the course of testicular damage and eventually affect fertility. Theoretically, attributed to the hypercoagulable state of vasculitis in patients with COVID-19, the testicular damage could be result of testicular segmental vascularization⁸⁴. One study has shown evidence of direct SARS-COV-2 infection of endothelial cells and diffuse endothelial inflammation⁸⁵, Endothelial dysfunction may be subsequent to organ ischemia⁸⁶, which might provide a rationale for one study that described ischemia-related priapism in a patient with COVID-19⁸⁷, suggesting that vasculitis-orchitis might have a crucial role in the development of the testicular injury caused by SARS-CoV-2 infection. Moreover, the intrusion of CD68+

macrophages into the interstitial tissue of the testes could contribute to a decline in steroidogenesis and testosterone⁶⁶, and the change in the hormonal profile might contribute to susceptibility to SARS-CoV-2 infection, leading to a more profound pathophysiological role in COVID-19 patients ⁸⁸.

During the SARS-CoV-2 outbreak, SARS-CoV-2 infected-cats also presented a profile of testicular atrophy ⁸⁹, and were reported to have acquired the infection from humans ⁹⁰. Furthermore, studies on chickens infected with coronavirus IBV also showed that immune cells infiltrated into the interstitium of the testis, which was responsible for the reduced fertility ^{25,91}.

In summary, the coronavirus-induced adaptive immune response might lead to testicular damage and endocrine abnormality, eventually disrupting spermatogenesis in patients recovering from COVID-19. However, this hypothesis remains to be confirmed and studies should be undertaken to establish an animal model to determine the underlying pathophysiological mechanisms and to mitigate the risk of testicular injury during COVID-19 disease. Precautions against SARS-CoV-2-induced male infertility should be taken.

3.3 Antibody-mediated infertility by SARS-CoV-2

In SARS-infected testes, Immunohistochemistry analysis showed a large amount of IgG precipitation in the seminiferous epithelium of the testis, as well as in degraded germ cells and Sertoli cells, suggesting that the extensive IgG triggered by a secondary autoimmune response might aggravate the testicular damage ²². In addition, deposits of IgG are associated with autoimmune orchitis (EAO)⁹², which might activate immune cells in the host to produce antibodies against the virus, as well as introducing antibodies into semen⁹³. In patients with

COVID-19, the positive rate of IgG reached 100%⁹⁴, especially antiphospholipid antibodies⁹⁵, which are antisperm antibodies that could interfere with fertilization⁹⁶, suggesting that male patients with COVID-19 should be cautioned against the adverse effects of a high IgG titer on their reproduction ability.

In healthy testis tissue, immune cells and cytokines are beneficial for the development of spermatogonia. However, the immune imbalance associated with infection and inflammation can contribute to male sterility. Overall, in addition to the pathogenic effects of coronavirus, the host-induced immune response against the virus also plays an important role in the overall disease process.

3.4 High fever and steroid-mediated infertility

It is generally believed that high fever can be detrimental to the normal function of the testes. Fever is one of the notable features of COVID-19³, and thus might play an important role in testicular dysfunction. Germ cells can develop at a normal pace at temperatures less than 37.8 °C; however, higher temperatures might cause irreversible damage to germ cells. Research confirms that high temperatures can lead to the meiotic arrest of germ cells⁹⁷. In general, increased body temperature has a negative influence on spermatogenesis and may ultimately lead to male infertility.

In addition, patients with COVID-19 were almost all affected by SARS-CoV-2-related stress and were advised to use steroids (methylprednisolone, 1–2 mg/kg per day)² to treat SARS-CoV-2; however, the stress⁹⁸ and corticosteroid therapy might have adverse effects on sexual function, such as reduced libido and erectile dysfunction^{99,100}. Leydig cells were also proven to be dysfunctional in glucocorticoid-treated rats¹⁰¹. Therefore, these observations

suggest that the assessment of fertility in patients with COVID-19 is imperative.

4. Conclusion

This review obtained clues from basic research on other viruses to understand how the novel SARS-CoV-2 virus might generate pathogenic effects in male fertility. We highlighted that male fertility might be highly vulnerable to SARS-CoV-2 infection. Infection with this novel virus not only seriously threatens an individual's overall health, but also might lead to male infertility. Perspectives gained from multi-organ research during the recent epidemic raises the possibility that damage to the male reproductive tract might be an underappreciated result of SARS-CoV-2 infection. Therefore, more attention should be paid to the effects on male fertility of SARS-CoV-2 infection, and should this causal link between SARS-CoV-2 infection and male infertility be confirmed, male patients should consider cryopreserving their sperm to preserve fertility.

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Authors' roles

Wenbing Zhu and Chuang Huang conceived and designed the study. Chuang Huang and Xiren Ji drafted the manuscript. Wenjun Zhou, Zhenghui Huang, Xiangjie Peng, Liqing Fan, and Ge Lin revised the drafts. All authors approved the final version of the manuscript.

Conflict of interest

None declared.

References

1. Zhu N, Zhang D, Wang W, et al. A Novel Coronavirus from Patients with Pneumonia in China, 2019. *N Engl J Med*. 2020;382(8):727-733.
2. Chen N, Zhou M, Dong X, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. *Lancet*. 2020;395(10223): 507-513.

- 337 3. Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus in
338 Wuhan, China. *The Lancet*. 2020;395(10223):497-506.
- 339 4. Li Q, Guan X, Wu P, et al. Early Transmission Dynamics in Wuhan, China, of Novel Coronavirus-
340 Infected Pneumonia. *the new england journal of medicine*. 2020;382(13):1199-1207.
- 341 5. Hoffmann M, Kleine-Weber H, Schroeder S, et al. SARS-CoV-2 Cell Entry Depends on ACE2 and
342 TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. *cell*. 2020;181(2):405-418.
- 343 6. Wang Z, Xu X. scRNA-seq Profiling of Human Testes Reveals the Presence of the ACE2 Receptor,
344 A Target for SARS-CoV-2 Infection in Spermatogonia, Leydig and Sertoli Cells. *cells*. 2020;9(4):
345 920-920.
- 346 7. Salam AP, Horby PW. The Breadth of Viruses in Human Semen. *emerging infectious diseases*.
347 2017;23(11):1922-1924.a
- 348 8. Klimova RR, Chichev EV, Naumenko VA, et al. Herpes simplex virus and cytomegalovirus in male
349 ejaculate: herpes simplex virus is more frequently encountered in idiopathic infertility and
350 correlates with the reduction in sperm parameters. *voprosy virusologii*. 2010;55(1):27-31.
- 351 9. Garrido N, Meseguer M, Remohi J, Simon C, Pellicer A. Semen characteristics in human
352 immunodeficiency virus (HIV)- and hepatitis C (HCV)-seropositive males: predictors of the success
353 of viral removal after sperm washing. *Hum Reprod*. 2005;20(4):1028-1034.
- 354 10. Paoli D, Pallotti F, Turriziani O, et al. SARS-CoV-2 presence in seminal fluid: Myth or reality.
355 *Journal of Andrology*. 2020.
- 356 11. Pan F, Xiao X, Guo J, et al. No evidence of severe acute respiratory syndrome-coronavirus 2 in
357 semen of males recovering from coronavirus disease 2019. *fertility and sterility*. 2020;113(6):1135-
358 1139.
- 359 12. Paoli D, Pallotti F, Colangelo S, et al. Study of SARS-CoV-2 in semen and urine samples of a
360 volunteer with positive naso-pharyngeal swab. *journal of endocrinological investigation*. 2020: 1-
361 4.
- 362 13. Song C, Wang Y, Li W, et al. absence of 2019 novel coronavirus in semen and testes of COVID-19
363 patients. *biology of reproduction*. 2020;103(1):4-6.
- 364 14. Li D, Jin M, Bao P, Zhao W, Zhang S. Clinical Characteristics and Results of Semen Tests Among
365 Men With Coronavirus Disease 2019. *JAMA Netw Open*. 2020;3(5):e208292.
- 366 15. Yang M, Chen S, Huang B, et al. Pathological Findings in the Testes of COVID-19 Patients: Clinical
367 Implications. *european urology focus*. 2020;6(5):1124-1129.
- 368 16. Liu Y, Yan LM, Wan L, et al. Viral dynamics in mild and severe cases of COVID-19. *lancet infectious
369 diseases*. 2020;20(6):656-657.
- 370 17. Rastrelli G, Di Stasi V, Inglese F, et al. Low testosterone levels predict clinical adverse outcomes in
371 SARS-CoV-2 pneumonia patients. *Andrology*. 2020.
- 372 18. Wang Z, Xu X. scRNA-seq Profiling of Human Testes Reveals the Presence of the ACE2 Receptor,
373 A Target for SARS-CoV-2 Infection in Spermatogonia, Leydig and Sertoli Cells. *Cells*. 2020;9(4):
374 920.
- 375 19. Kelly JC, Dombrowski M, O'Neil-Callahan M, Kernberg AS, Frolova AI, Stout MJ. False-Negative
376 COVID-19 Testing: Considerations in Obstetrical Care. In:2020.
- 377 20. Li W, Shi Z, Yu M, et al. Bats are natural reservoirs of SARS-like coronaviruses. *Science*. 2005;310
378 (5748):676-679.
- 379 21. Kupferschmidt K. Emerging diseases. Researchers scramble to understand camel connection to
380 MERS. *Science*. 2013;341(6147):702.

- 381 22. Xu J, Qi L, Chi X, et al. Orchitis: a complication of severe acute respiratory syndrome (SARS). *Biol*
382 *Reprod.* 2006;74(2):410-416.
- 383 23. Shastri A, Wheat J, Agrawal S, et al. Delayed clearance of SARS-CoV2 in male compared to female
384 patients: High ACE2 expression in testes suggests possible existence of gender-specific viral
385 reservoirs. *MedRxiv*, In:2020.
- 386 24. Sigurdardóttir ÓG, Kolbjørnsen, Lutz HJJCP. Orchitis in a Cat Associated with Coronavirus
387 Infection. 2001;124(2-3):219-222.
- 388 25. Boltz DA, Zimmerman CR, Nakai M, Bunick D, Scherba G, Bahr JMJAd. Epididymal stone formation
389 and decreased sperm production in roosters vaccinated with a killed strain of avian infectious
390 bronchitis virus. 2006;50(4):594-598.
- 391 26. Situ J, Wang W, Long F, et al. Hepatitis E viral infection causes testicular damage in mice. *Virology*.
392 2020;541:150-159.
- 393 27. Govero J, Esakky P, Scheaffer SM, et al. Zika virus infection damages the testes in mice. *Nature*.
394 2016;540(7633):438-442.
- 395 28. Ma W, Li S, Ma S, et al. Zika Virus Causes Testis Damage and Leads to Male Infertility in Mice. *Cell*.
396 2017;168(3):542.
- 397 29. Li W, Moore MJ, Vasilieva N, et al. Angiotensin-converting enzyme 2 is a functional receptor for
398 the SARS coronavirus. *Nature*. 2003;426(6965):450-454.
- 399 30. Hoffmann M, Kleine-Weber H, Krüger N, Müller M, Drosten C, Pöhlmann S. The novel coronavirus
400 2019 (2019-nCoV) uses the SARS-coronavirus receptor ACE2 and the cellular protease TMPRSS2
401 for entry into target cells. *bioRxiv*. 2020:2020.2001.2031.929042.
- 402 31. Zhou P, Yang X-L, Wang X-G, et al. A pneumonia outbreak associated with a new coronavirus of
403 probable bat origin. *Nature*. 2020;579(7798):270-273.
- 404 32. Tipnis SR, Hooper NM, Hyde R, Karran E, Christie G, Turner AJ. A Human Homolog of Angiotensin
405 -converting Enzyme CLONING AND FUNCTIONAL EXPRESSION AS A CAPTOPRIL-INSENSITIVE
406 CARBOXYPEPTIDASE. *journal of biological chemistry*. 2000;275(43): 33238-33243.
- 407 33. Harmer D, Gilbert M, Borman R, Clark KL. Quantitative mRNA expression profiling of ACE 2, a
408 novel homologue of angiotensin converting enzyme. *febs letters*. 2002;532(1):107-110.
- 409 34. Douglas GC, O'Bryan MK, Hedger MP, et al. The novel angiotensin-converting enzyme (ACE)
410 homolog, ACE2, is selectively expressed by adult Leydig cells of the testis. *endocrinology*. 2004;
411 145(10):4703-4711.
- 412 35. Corona G, Baldi E, Isidori AM, et al. SARS-CoV-2 infection, male fertility and sperm
413 cryopreservation: a position statement of the Italian Society of Andrology and Sexual Medicine
414 (SIAMS) (Societa Italiana di Andrologia e Medicina della Sessualita). *J Endocrinol Invest*. 2020;43(8):
415 1153-1157.
- 416 36. Zhao J-m, Zhou G-d, Sun Y-l, et al. Clinical pathology and pathogenesis of severe acute
417 respiratory syndrome. *chinese journal of clinical hepatology*. 2003;17(3):217-221.
- 418 37. Puggioni G, Pintus D, Melzi E, et al. Testicular Degeneration and Infertility following Arbovirus
419 Infection. *J Virol*. 2018;92(19).
- 420 38. Shang J, Ye G, Shi K, et al. Structural basis of receptor recognition by SARS-CoV-2. *Nature*. 2020;
421 581(7807):221-224.
- 422 39. Perlman S, Netland J. Coronaviruses post-SARS: update on replication and pathogenesis. *nature*
423 *reviews microbiology*. 2009;7(6):439-450.
- 424 40. Stanley KE, Thomas E, Leaver M, Wells D. Coronavirus disease-19 and fertility: viral host entry

- protein expression in male and female reproductive tissues. *fertility and sterility*. 2020;114(1): 33-43.
41. Neto FT, Bach PV, Najari BB, Li PS, Goldstein M. Spermatogenesis in humans and its affecting factors. *Semin Cell Dev Biol*. 2016;59:10-26.
 42. Guan X, Chen F, Chen P, et al. Effects of spermatogenic cycle on Stem Leydig cell proliferation and differentiation. *Mol Cell Endocrinol*. 2019;481:35-43.
 43. Eroschenko VP, Wilson WO, Siopes TD. Function and histology of testes from aged coturnix maintained on different photoperiods. *J Gerontol*. 1977;32(3):279-285.
 44. Uraki R, Hwang J, Jurado KA, et al. Zika virus causes testicular atrophy. *science advances*. 2017;3(2): 1602899-1602899.
 45. Chen S-R, Liu Y-X. Regulation of spermatogonial stem cell self-renewal and spermatocyte meiosis by Sertoli cell signaling. *reproduction*. 2015;149(4).
 46. Anderson RA, Irvine DS, Balfour C, Groome NP, Riley SC. Inhibin B in seminal plasma: testicular origin and relationship to spermatogenesis. *Hum Reprod*. 1998;13(4):920-926.
 47. Mahmoud AM, Comhaire FH, Depuydt CE. The clinical and biologic significance of serum inhibins in subfertile men. *Reprod Toxicol*. 1998;12(6):591-599.
 48. Cavanagh D. Nidovirales: a new order comprising Coronaviridae and Arteriviridae. *Arch Virol*. 1997;142(3):629-633.
 49. Gallardo RA, Hoerr FJ, Berry WD, van Santen VL, Toro H. Infectious bronchitis virus in testicles and venereal transmission. *Avian Dis*. 2011;55(2):255-258.
 50. Jones RC. Europe: history, current situation and control measures for infectious bronchitis. *brazilian journal of poultry science*. 2010;12(2):125-128.
 51. Villarreal L, Brandão PE, Chacón JL, et al. Orchitis in roosters with reduced fertility associated with avian infectious bronchitis virus and avian metapneumovirus infections. 2007;51(4):900-904.
 52. Guan W-j, Ni Z-y, Hu Y, et al. Clinical Characteristics of Coronavirus Disease 2019 in China. *the new england journal of medicine*. 2020;382(18):1708-1720.
 53. Verma S, Saksena S, Sadri-Ardekani H. ACE2 receptor expression in testes: implications in coronavirus disease 2019 pathogenesis†. *biology of reproduction*. 2020.
 54. Aversa A, Jannini EA. COVID-19, or the triumph of monogamy? *minerva endocrinologica*. 2020; 45(2):77-78.
 55. Gallardo RA, Hoerr FJ, Berry WD, Santen VL, Toro H. Infectious Bronchitis Virus in Testicles and Venereal Transmission. *avian diseases*. 2011;55(2):255-258.
 56. Perry MJ, Arrington S, Neumann LM, Carrell D, Mores CN. It is currently unknown whether SARS-CoV-2 is viable in semen or whether COVID-19 damages spermatozoa. *Andrology*. 2020.
 57. Boltz DA, Nakai M, Bahra JM. Avian infectious bronchitis virus: a possible cause of reduced fertility in the rooster. *Avian Dis*. 2004;48(4):909-915.
 58. Holtmann N, Edimiris P, Andree M, et al. Assessment of SARS-CoV-2 in human semen—a cohort study. *Fertility and Sterility*. 2020;114(2):233-238.
 59. Benyeda Z, Mató T, Süveges T, et al. Comparison of the pathogenicity of QX-like, M41 and 793/B infectious bronchitis strains from different pathological conditions. *avian pathology*. 2009;38(6): 449-456.
 60. Boltz DA, Nakai M, Bahr JM. Avian Infectious Bronchitis Virus: A Possible Cause of Reduced Fertility in the Rooster. *avian diseases*. 2004;48(4):909-915.
 61. Villarreal LYB, Brandão PE, Chacón JLV, et al. Orchitis in Roosters with Reduced Fertility Associated

- with Avian Infectious Bronchitis Virus and Avian Metapneumovirus Infections. *avian diseases*. 2007; 51(4):900-904.
62. Liu Y, Ding Z. Obesity, a serious etiologic factor for male subfertility in modern society. *reproduction*. 2017;154(4).
 63. Li N, Wang T, Han D. Structural, cellular and molecular aspects of immune privilege in the testis. *frontiers in immunology*. 2012;3:152-152.
 64. Loveland KL, Klein B, Poeschl D, et al. Cytokines in Male Fertility and Reproductive Pathologies: Immunoregulation and Beyond. *frontiers in endocrinology*. 2017;8:307-307.
 65. Zhao S, Zhu W, Xue S, Han D. Testicular defense systems: immune privilege and innate immunity. *cellular & molecular immunology*. 2014;11(5):428-437.
 66. Hedger MP, Meinhardt A. Cytokines and the immune-testicular axis. *Journal of Reproductive Immunology*. 2003;58(1):1-26.
 67. Mruk DD, Cheng CY. The Mammalian Blood-Testis Barrier: Its Biology and Regulation. *Endocr Rev*. 2015;36(5):564-591.
 68. Liu Q, Wang RS, Qu GQ, et al. Gross examination report of a COVID-19 death autopsy. *Fa Yi Xue Za Zhi*. 2020;36(1):21-23.
 69. Pedersen SF, Ho Y-C. SARS-CoV-2: a storm is raging. *journal of clinical investigation*. 2020;130(5):2202-2205.
 70. Lui W-Y, Lee WM, Cheng CY. Transforming Growth Factor- β 3 Perturbs the Inter-Sertoli Tight Junction Permeability Barrier in Vitro Possibly Mediated via Its Effects on Occludin, Zonula Occludens-1, and Claudin-11. *endocrinology*. 2001;142(5):1865-1877.
 71. Dobashi M, Fujisawa M, Yamazaki T, Okada H, Kamidono S. Distribution of intracellular and extracellular expression of transforming growth factor-beta1 (TGF-beta1) in human testis and their association with spermatogenesis. *asian journal of andrology*. 2002;4(2):105-109.
 72. Itman C, Mendis SHS, Barakat BM, Loveland KAL. All in the family: TGF- β family action in testis development. *reproduction*. 2006;132(2):233-246.
 73. Maiorino MI, Bellastella G, Giugliano D, Esposito K. From inflammation to sexual dysfunctions: a journey through diabetes, obesity, and metabolic syndrome. *journal of endocrinological investigation*. 2018;41(11):1249-1258.
 74. Ochsenkühn R, O'Connor AE, Hirst JJ, Baker HWG, Kretser DMd, Hedger MP. The relationship between immunosuppressive activity and immunoregulatory cytokines in seminal plasma: Influence of sperm autoimmunity and seminal leukocytes. *journal of reproductive immunology*. 2006;71(1):57-74.
 75. Hales DB. Interleukin-1 inhibits Leydig cell steroidogenesis primarily by decreasing 17 alpha-hydroxylase/C17-20 lyase cytochrome P450 expression. *endocrinology*. 1992;131(5):2165-2172.
 76. Janssen SJ, Kirby JD, Hess RA, et al. Identification of epididymal stones in diverse rooster populations. *poultry science*. 2000;79(4):568-574.
 77. Mahecha GAB, Oliveira CA, Balzuweit K, Hess RA. Epididymal lithiasis in roosters and efferent ductule and testicular damage. *reproduction*. 2002;124(6):821-834.
 78. Pozzilli P, Lenzi A. Commentary: Testosterone, a key hormone in the context of COVID-19 pandemic. *Metabolism*. 2020;108:154252.
 79. Wambier CG, Goren A. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection is likely to be androgen mediated. *journal of the american academy of dermatology*. 2020;83(1):308-309.

- 513 80. S W, Q Y, S F, et al. Characteristics of lymphocyte subsets and cytokines in peripheral blood of 123
514 hospitalized patients with 2019 novel coronavirus pneumonia (NCP). In:2020.
- 515 81. Liu J, Li S, Liu J, et al. Longitudinal characteristics of lymphocyte responses and cytokine profiles in
516 the peripheral blood of SARS-CoV-2 infected patients. *ebiomedicine*. 2020;102763-102763.
- 517 82. Zhang H, Yin Y, Wang G, Liu Z, Liu L, Sun F. Interleukin-6 disrupts blood-testis barrier through
518 inhibiting protein degradation or activating phosphorylated ERK in Sertoli cells. *scientific reports*.
519 2015;4(1):4260-4260.
- 520 83. Xu J, Qi L, Chi X, et al. Orchitis: a complication of severe acute respiratory syndrome (SARS). 2006;
521 74(2):410-416.
- 522 84. Leisman DE, Deutschman CS, Legrand M. Facing COVID-19 in the ICU: vascular dysfunction,
523 thrombosis, and dysregulated inflammation. *intensive care medicine*. 2020;46(6):1105-1108.
- 524 85. Varga Z, Flammer AJ, Steiger P, et al. Endothelial cell infection and endotheliitis in COVID-19. *the*
525 *lancet*. 2020;395(10234):1417-1418.
- 526 86. Bonetti PO, Lerman LO, Lerman A. Endothelial Dysfunction A Marker of Atherosclerotic Risk.
527 *arteriosclerosis thrombosis and vascular biology*. 2003;23(2):168-175.
- 528 87. Lamamri M, Chebbi A, Mamane J, et al. Priapism in a patient with coronavirus disease 2019
529 (COVID-19): A case report. *Am J Emerg Med*. 2020.
- 530 88. Salonia A, Corona G, Giwercman A, et al. SARS-CoV-2, testosterone and frailty in males
531 (PROTEGGIMI): A multidimensional research project. *Andrology*. 2020.
- 532 89. Sigurdardottir OG, Kolbjornsen O, Lutz H. Orchitis in a cat associated with coronavirus infection. *J*
533 *Comp Pathol*. 2001;124(2-3):219-222.
- 534 90. Zhang Q, Zhang H, Huang K, et al. SARS-CoV-2 neutralizing serum antibodies in cats: a
535 serological investigation. In:2020.
- 536 91. Benyeda Z, Szeredi L, Mató T, et al. Comparative histopathology and immunohistochemistry of
537 QX-like, Massachusetts and 793/B serotypes of infectious bronchitis virus infection in chickens.
538 *journal of comparative pathology*. 2010;143(4):276-283.
- 539 92. Itoh M, Hiramane C, Tokunaga Y, Mukasa A, Hojo K. A new murine model of autoimmune orchitis
540 induced by immunization with viable syngeneic testicular germ cells alone. II.
541 Immunohistochemical findings of fully-developed inflammatory lesion. *autoimmunity*. 1991;10(2):
542 89-97.
- 543 93. Moldoveanu Z, Huang WQ, Kulhavy R, Pate MS, Mestecky J. Human male genital tract secretions:
544 both mucosal and systemic immune compartments contribute to the humoral immunity. *J*
545 *Immunol*. 2005;175(6):4127-4136.
- 546 94. Long QX, Liu BZ, Deng HJ, et al. Antibody responses to SARS-CoV-2 in patients with COVID-19.
547 *nature medicine*. 2020;26(6):845-848.
- 548 95. Connell NT, Battinelli EM, Connors JM. Coagulopathy of COVID-19 and antiphospholipid
549 antibodies. *journal of thrombosis and haemostasis*. 2020.
- 550 96. Chiu WWC, Chamley LW. Clinical associations and mechanisms of action of antisperm antibodies.
551 *fertility and sterility*. 2004;82(3):529-535.
- 552 97. Xu J, Xu Z, Jiang Y, Qian X, Huang Y. Cryptorchidism induces mouse testicular germ cell apoptosis
553 and changes in bcl-2 and bax protein expression. *journal of environmental pathology toxicology*
554 *and oncology*. 2000;19:25-33.
- 555 98. Chan JC, Morgan CP, Leu NA, et al. Reproductive tract extracellular vesicles are sufficient to
556 transmit intergenerational stress and program neurodevelopment. *nature communications*.

2020;11(1).

99. Ln C, Am M, Mm D, et al. Glucocorticoids: their role on gonadal function and LH secretion. *minerva endocrinologica*. 1996;21(2).
100. Scaroni C, Favia G, Lumachi F, et al. Unilateral adrenal tumor, erectile dysfunction and infertility in a patient with 21-hydroxylase deficiency: effects of glucocorticoid treatment and surgery. *experimental and clinical endocrinology & diabetes*. 2003;111(1):41-43.
101. Gao H-B, Tong M-H, Hu Y-Q, Guo Q-S, Ge R, Hardy MP. Glucocorticoid Induces Apoptosis in Rat Leydig Cells. *endocrinology*. 2002;143(1):130-138.